Association between Fusobacterium nucleatum and Colorectal Cancer (CRC)

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Outline of Content

A) Brief on human microbiota and cancer.

- Human Microbiome
- What is microbiota?
- Microbiota-related diseases
- Mechanism of microbiome carcinogenesis



- B) Fusobacterium nucleatum (F. nucleatum) and colorectal cancer (CRC)
- Microbiota in CRC
- Fusobacteria as biomarker in CRC
- Mechanism and immunity

Detection and therapeutic planning
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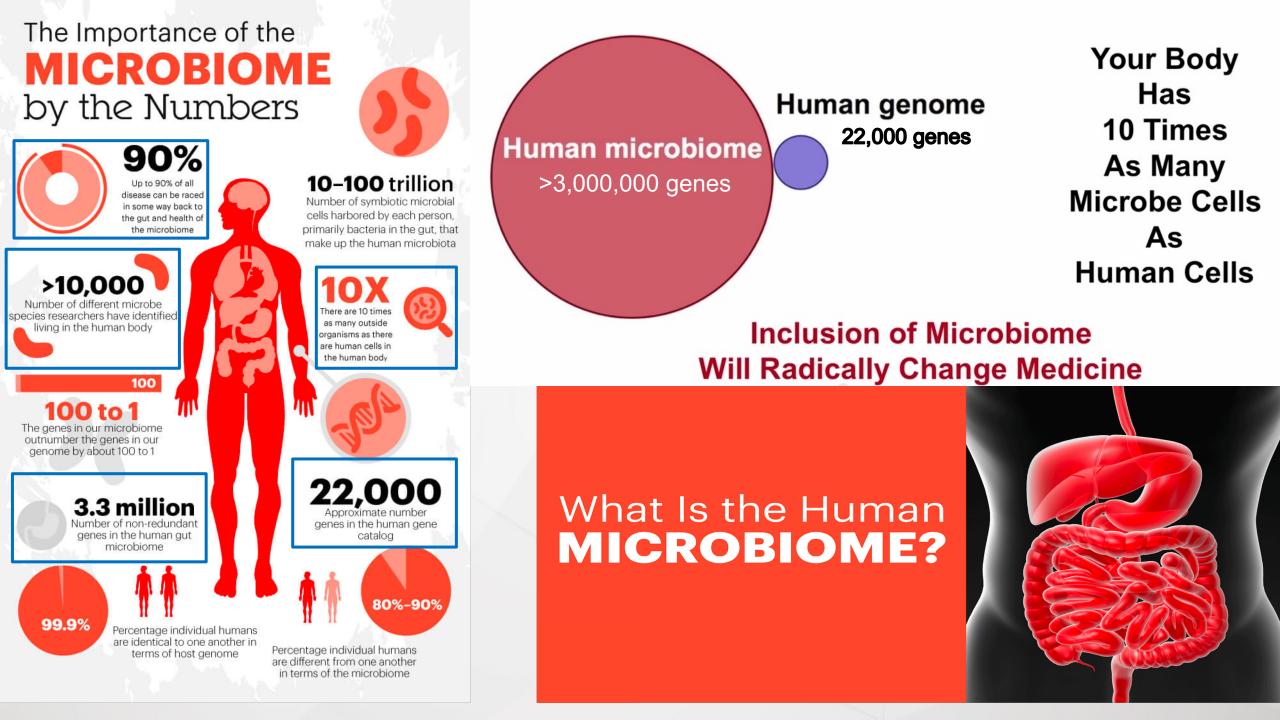


Brief on human microbiota and cancer

Fusobacterium and Colorectal Cancer

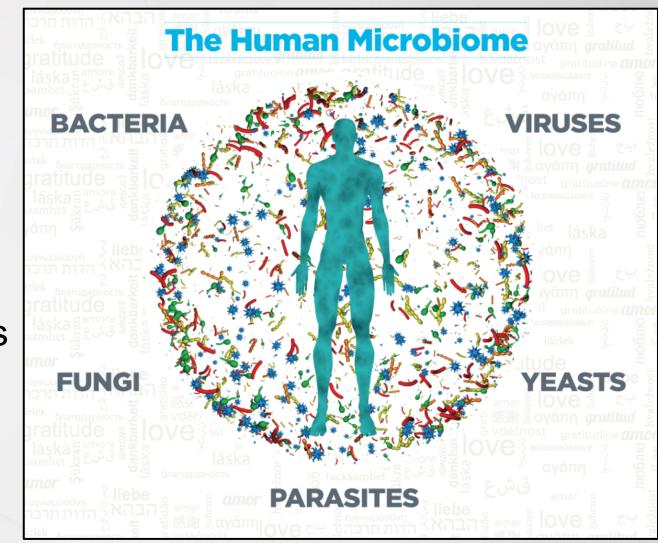
(CRC)

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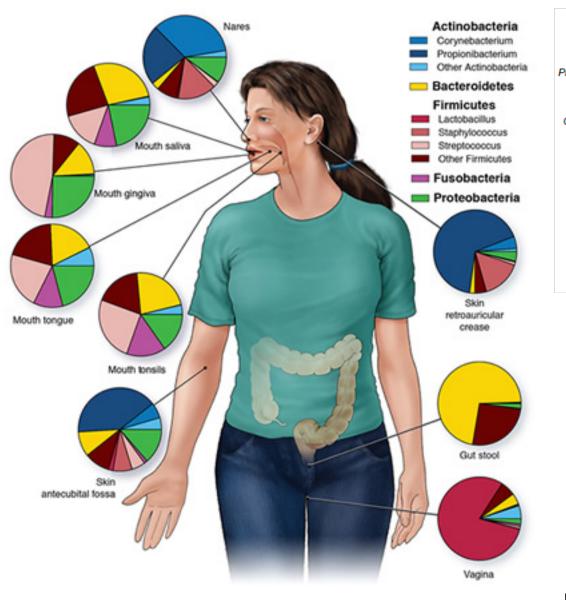


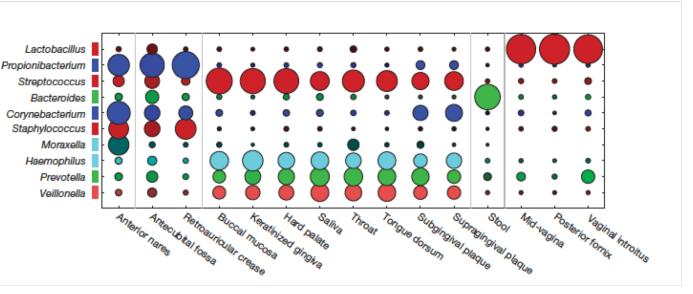
What is microbiota?

- Microbiota is an ecological community of microorganisms.
- bacteria, viruses, yeasts, parasites and fungi.
- immunologic, hormonal and metabolic homeostasis of hosts
- Potential carcinogenesis



Abundance of human microbiota

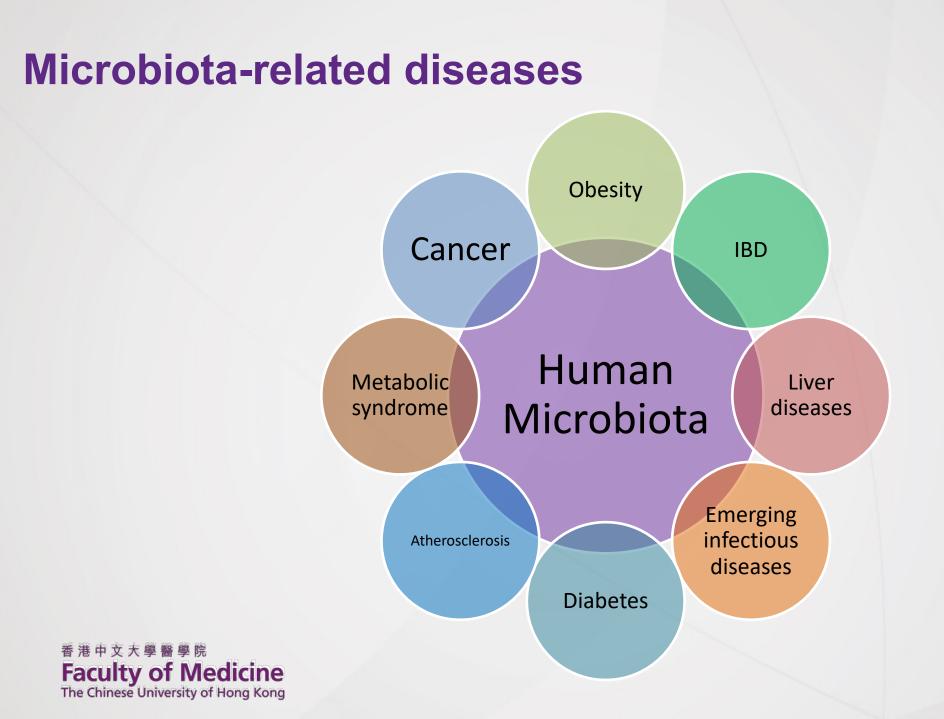




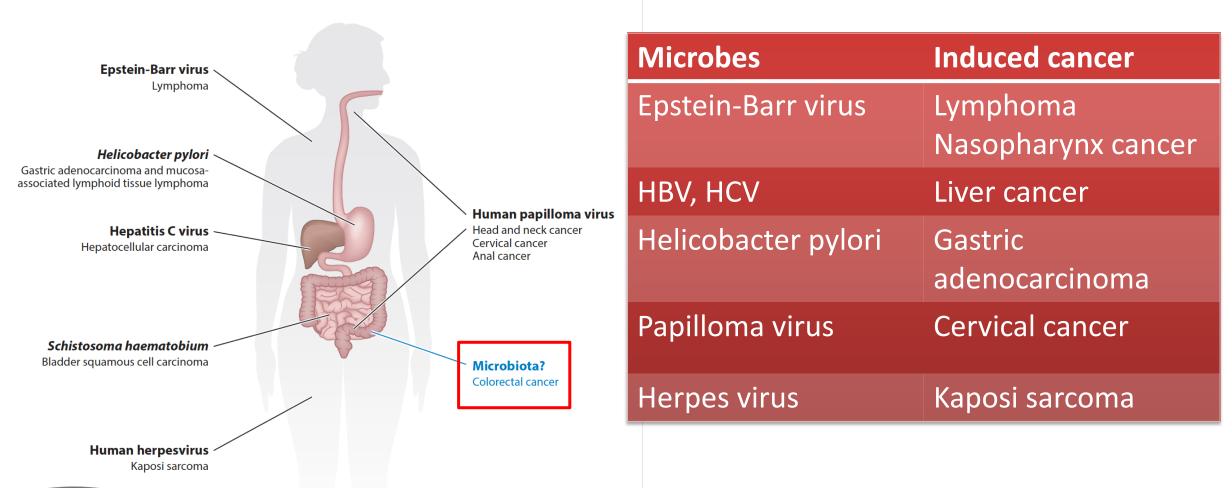
- The microbiome of each organ is distinct
- The effects on diseases are organ specific



Human Microbiome Project, C. (2012). "Structure, function and diversity of the healthy human microbiome." <u>Nature</u> **486**(7402): 207-214



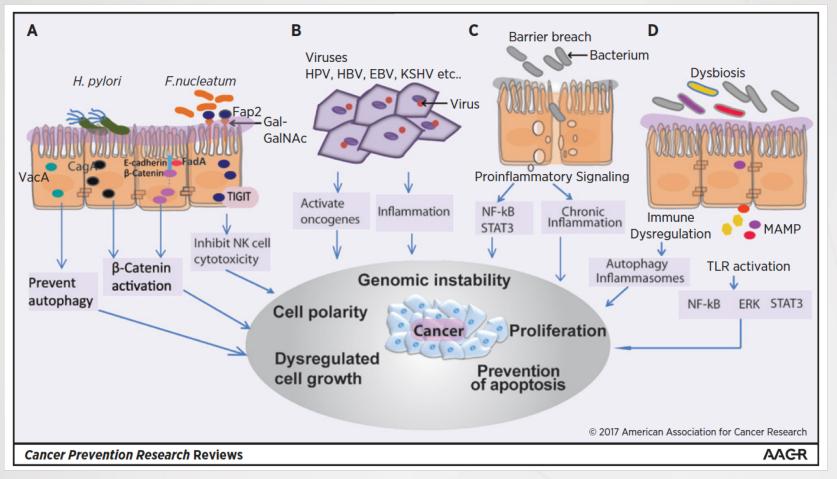
Cancer Contributor





Brennan, C. A. and W. S. Garrett (2016). "Gut Microbiota, Inflammation, and Colorectal Cancer." <u>Annu Rev Microbiol</u> **70**: 395-411.

Mechanisms of microbiome carcinogenesis



- Microbes inject **effectors** into the host cells.
- **Oncoproteins** of tumor virus transform cells types.
- **Barrier breach** results in proinflammatory signaling for carcinogenesis.
- **Dysbiosis** and altered microbiota-host interaction can induce carcinogenesis.



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Rajagopala, S. V., et al. (2017). "The Human Microbiome and Cancer." Cancer Prev Res (Phila) 10(4): 226-234.

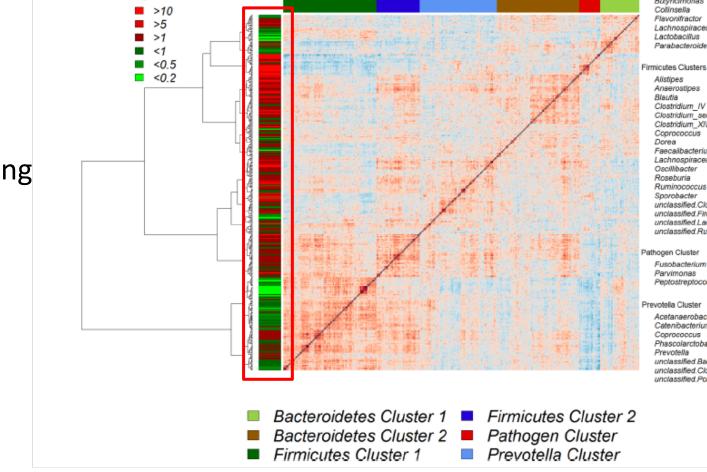
Brief on human microbiota and cancer

Fusobacterium and Colorectal Cancer (CRC)

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Microbiota in CRC

- The abundance of diverse • microbes is different in CRC tissue and normal tissue.
- There is correlation among ۲ microbes.
- The dysbiosis cause the ۲ CRC.



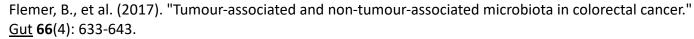
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Color Key

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4.5

Fold change CRC vs HC



Bacteroidetes Clusters Acetanaerobacterium Alistipes

Bacteroides

Bilophila Butyricimonas

Collinsella

Flavonifractor

Lactobacillus Parabacteroides

Alistipes Anaerostipes Blautia Clostridium_IV

Oscillibacter Roseburia Ruminococcus Sporobacter unclassified, Clostridiales

Lachnospiracea /S

Clostridium_sensu_stricto Clostridium_XIVa Coprococcus Dorea Faecalibacterium Lachnospiracea IS

unclassified.Firmicutes unclassified.Lachnospiraceae unclassified.Ruminococcaceae

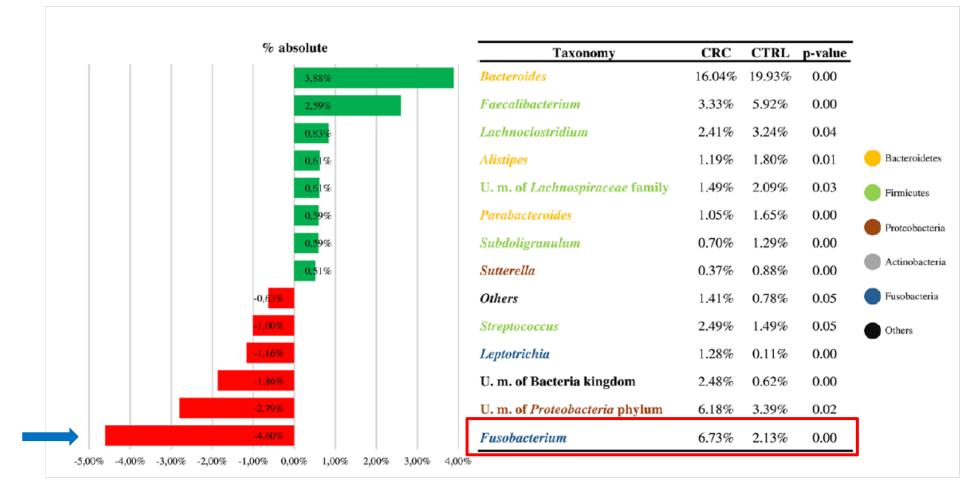
Fusobacterium Parvimonas Peptostreptococcus

Acetanaerobacterium Catenibacterium Coprococcus Phascolarctobacterium Prevotella

unclassified.Bacteroidetes unclassified.Clostridiales unclassified,Porphyromonadaceae

Bifidobacterium

Microbiota in CRC

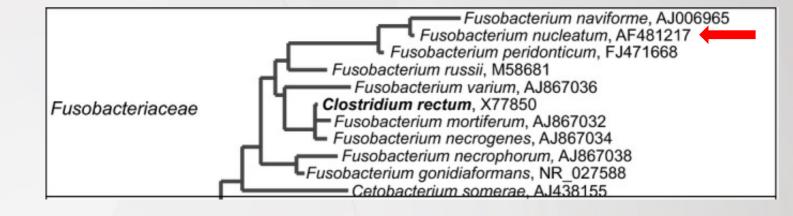


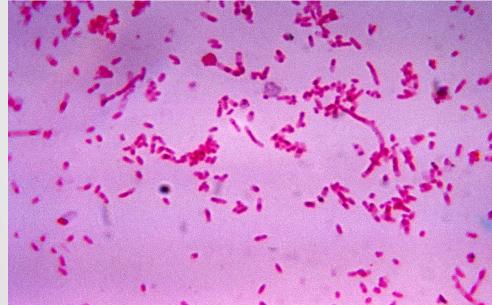
- Fusobacterium is the key phylotypes that contribute to the dysbiosis in CRC patients.
- Compared to health tissue, Fusobacterium is more abundant in CRC tissue.

Mancabelli, L., et al. (2017). "Identification of universal gut microbial biomarkers of common human intestinal diseases by metaanalysis." <u>FEMS Microbiol Ecol</u> **93**(12).

Introduction of Fusobacteria

- Gram-negative anaerobic bacterium.
- Pleomorphic, but usually spindle-shaped.





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- Over 30 species within *Fusobacterium*.
- *F. nucleatum* is mostly linked to oral diseases.

Fusobacterium as the biomarker in CRC

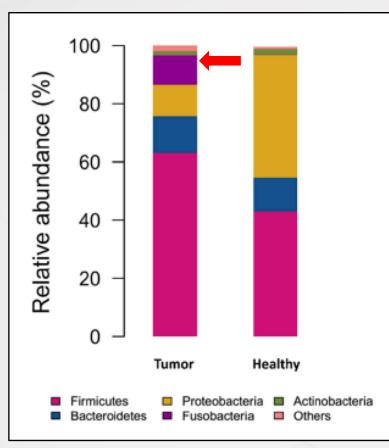


Table 2. Association between *Fusobacterium* abundance andcolorectal adenomas.

Categories*		Case (n = 48)	Control (n = 67)	OR (95% CI)**
Tertile 1	low	8	23	Reference
Tertile 2		12	22	1.57 (0.54-4.57)
Tertile 3	high	28	22	3.66 (1.37–9.74)
P trend				

*The abundance of *Fusobacterium* among control subjects were used to generate tertile cut-off. The lowest tertile of *Fusobacterium* abundance was considered as the reference.

**Odds ratio and 95% confidence interval.

Compared to subjects with a low copy number, subjects with high abundance of *Fusobacterium* are more likely to be adenoma cases than controls.

• *Fusobacterium* is significantly more abundant in CRC tissue than normal tissue.

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McCoy, A. N., et al. (2013). "Fusobacterium is associated with colorectal adenomas." <u>PLoS One</u> **8**(1): e53653

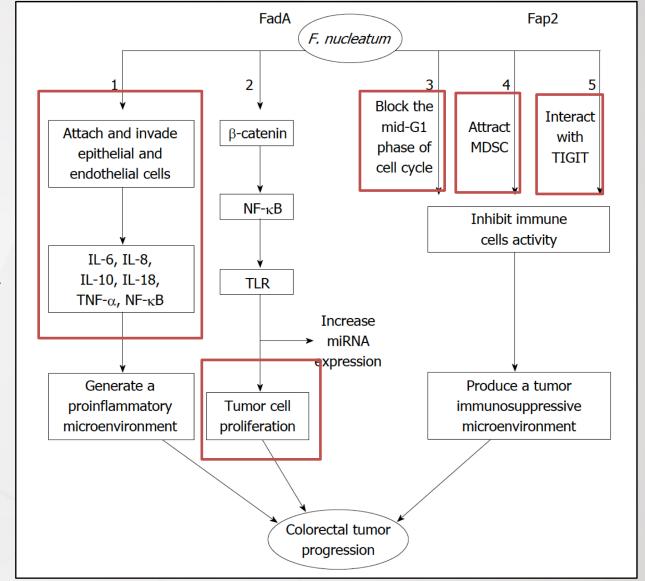
Mechanism of F. nucleatum carcinogenesis

FadA (Fusobacterium adhesion A)

- Induce cytokines to generate a proinflammatory microenvironment.
- Activate the β-catenin signaling pathway to promote tumor cell proliferation.

Fap2 (fibroblast activation protein 2)

- Block the G1 phase of cell cycle.
- Attract MDSC to promote tumor progression.
- Interaction with TIGIT to protect tumor cell from immune cell attack.



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Shang, F. M. and H. L. Liu (2018). "Fusobacterium nucleatum and colorectal cancer: A review." <u>World J Gastrointest Oncol</u> **10**(3): 71-81.

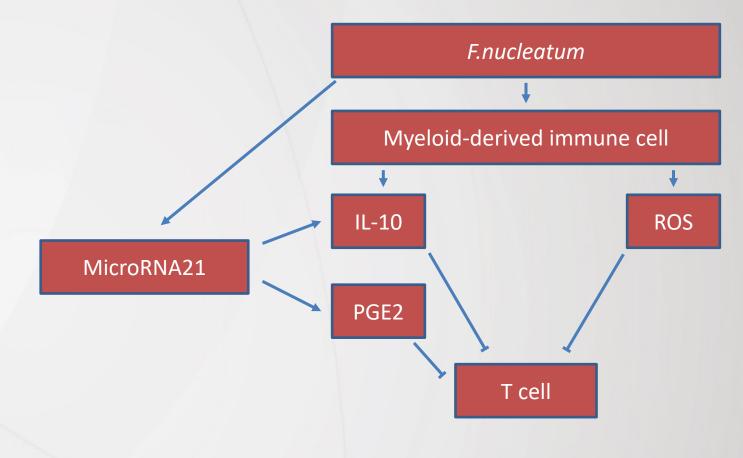
Immunity in F. nucleatum infection

- Myeloid-derived immune cells were enriched in *F. nucleatum* infected host.
- IL-10 and ROS were accumulated by myeloid-derived immune cell.
- Increased MicroRNA21 promote the expression of PGE2
- IL-10,ROS and PGE2 suppressively modulated T-cell-mediated adaptive immunity

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Nosho, K., et al. (2016). "Association of Fusobacterium nucleatum with immunity and molecular alterations in colorectal cancer." <u>World J Gastroenterol</u> **22**(2): 557-566.

Fusobacteria associated diseases

Anatomic sites	Disease		
Oral infection	Aggressive periodontitis		
	Chronic periodontitis		
	Endodontic infections		
Adverse pregnancy outcomes	Chorioamnionitis		
	Neonatal sepsis		
GI disorders	Colorectal cancer		
	Appendicitis		
Other infections	Cerebral aneurysm		
	Alzheimer's disease		
	Lemierre's syndrome		

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Han, Y. W. (2015). "Fusobacterium nucleatum: a commensal-turned pathogen." <u>Curr Opin Microbiol</u> 23: 141-147.

Detection of F. nucleatum in CRC

Total cases (n)	Positive cases (n)	Positive percentage	Detection method	Detection samples
101	88	87.13%	FISH and FQ-PCR	Frozen tissue and FFPE tissue
598	76	13%	qPCR	FFPE tissue
511	44	8.6%	qPCR	FFPE tissue
149	111	74%	qPCR	Genomic DNA
511	286	56%	qPCR	FFPE tissue
158	85	54%	ddPCR	Feces

- <u>METHOD</u> --- FISH (Fluorescence *in situ* hybridization), FQ-PCR, qPCR and ddPCR are the usual method in clinical detection.
- **<u>SAMPLE</u>** --- Feces are difficult to detect *F. nucleatum*, FFPE (Formalin-fixed paraffin embedded) tissues and frozen tissue are limited by surgery or colonoscopy.
- **<u>CONPARISON</u>** --- qPCR is most usual in detection, ddPCR in higher detection rate of

low concentration of sample, FQ-PCR displays higher sensitivity and specificity.

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Therapeutic planning



Chemoprevention

- Use of Aspirin, COX-2 inhibitor, EO2 antagonist.
- Induces neutrophils apoptosis and lipoxin-driven

immune –regulatory effect

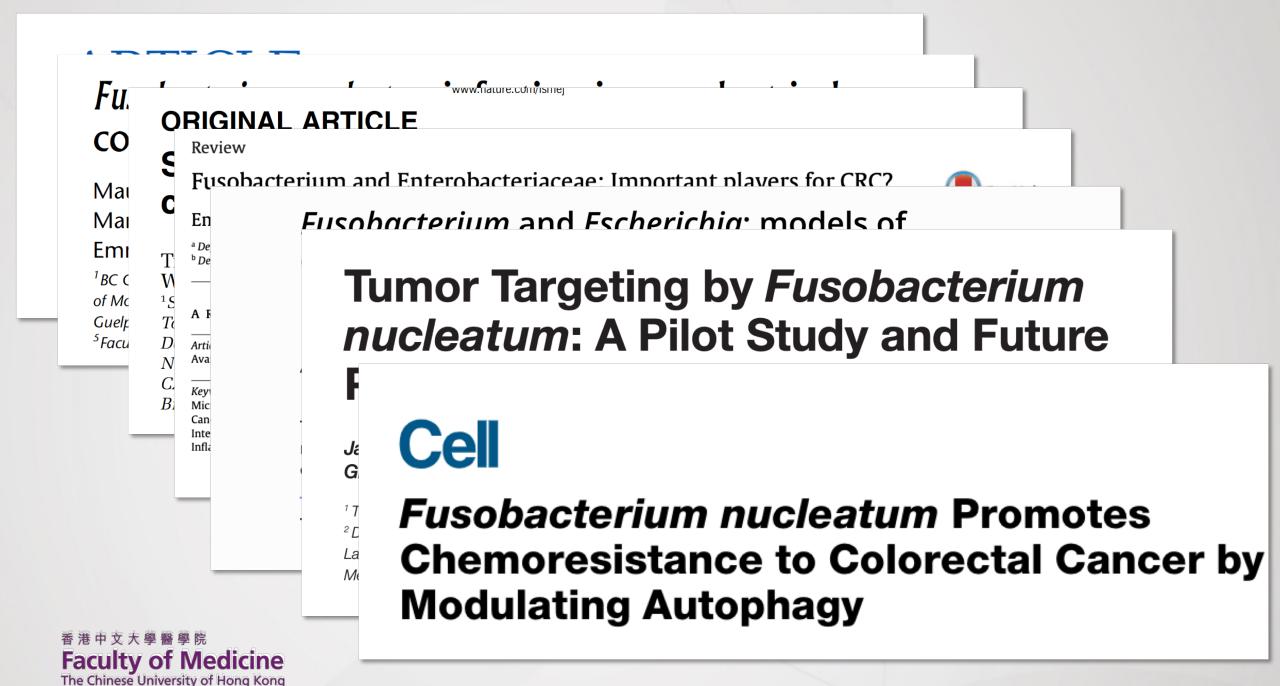
• Aspirin may support the immune system and prevent the development of *F. nucleatum*-associated CRC.



Immunotherapy

- Antibody treatment, immune-checkpoint blockade therapy, and adoptive cell transfer.
- Eg. 1) Anti-Fap2 antibody may favor antitumor immune response; 2) The blockade of CTLA-4 and PD-1 may shape the antitumor immune response.

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Summary

Gut microbiota has been extensively associated with

diverse cancers and diseases.

- *F. nucleatum* may contribute to CRC *via* multiple mechanisms.
- Chemoprevention and immunotherapy strategies could be most potential approaches to cure microbiota-related

Cancers. 香港中文大學醫學院 Faculty of Medicine The Chinese University of Hong Kong

Thanks for attention



References

Castellarin, M., et al. (2012). "Fusobacterium nucleatum infection is prevalent in human colorectal carcinoma." Genome Res 22(2): 299-306.

Rajagopala, S. V., et al. (2017). "The Human Microbiome and Cancer." <u>Cancer Prev Res (Phila)</u> 10(4): 226-234.

Wang, T., et al. (2012). "Structural segregation of gut microbiota between colorectal cancer patients and healthy volunteers." <u>ISME J</u> 6(2): 320-329

McCoy, A. N., et al. (2013). "Fusobacterium is associated with colorectal adenomas." PLoS One 8(1): e53653

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